# The syndrome "basses richesses" of sugar beet in France is associated with different pathogen types and insect vectors

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#### **Abstract**

The syndrome "basses richesses" (SBR) of sugar beet in France is associated with two phloem-restricted uncultivable bacteria: a stolbur phytoplasma and a γ-3 proteobacteria. The known vector of proteobacteria is a cixiid planthopper, *Pentastiridius leporinus* (Hemiptera Cixiidae), formerly shown to transmit both the prokaryotes. The role of *P. leporinus* and of two other planthopper species, *Cixius wagneri* and *Hyalesthes obsoletus*, in spreading the two pathogens to sugar beet were compared and quantified. Because of its abundance and high infection rates with proteobacterium, *P. leporinus* was confirmed to be the economic vector of SBR disease. *P. leporinus* and *C. wagneri* were infected by and transmitted proteobacterium, neither was infected by the phytoplasma. Neither of two populations of *H. obsoletus* living near sugar beet fields on bindweed and nettle respectively, carried proteobacteriumbut they were highly infected with two RFLP-differentiable stolbur types. Only the bindweed stolbur type was transmitted and pathogenic to sugar beets. Symptoms associated with the two prokaryotes were similar, but stolbur caused a stronger reduction on taproot biomass and sugar content than proteobacteria. This work underlines possible confusion in aetiology of diseases associated with phytoplasmas or phloem-restricted proteobacteria and the increasing importance of the latter plant pathogens.

**Key words:** syndrome "basses richesses", stolbur phytoplasma, phloem-restricted proteobacteria, planthoppers, cixiids, *Hyalesthes obsoletus, Cixius wagneri, Pentastiridius leporinus*.

#### Introduction

Hyalesthes obsoletus Signoet, Reptalus panzeri (Löw) Reptalus quinquecostatus (Dufour) and Pentastiridius sp., later identified as Pentastiridius leporinus (L.) (Nusillard, personal communication) are cixiid species known as vectors or suspected vectors of stolbur phytoplasmas (Stp) in Europe. Different Stp types have been associated with both different host plants and H. obsoletus populations (Langer and Maixner, 2004).

Other plant pathogenic phloem-restricted bacteria are proteobacteria. Marginal chlorosis of strawberry and syndrome "basses richesses" (SBR) of sugar beet are associated with two related γ-3 proteobacteria in the Arsenophonus clade, i.e 'Candidatus Phlomobacter fragariae' and SBR proteobacterium (SBRpr), transmitted respectively by Cixius wagneri China and P. leporinus (Danet et al., 2002; Gatineau et al., 2002; Sémétey et al., 2007b). SBR can also be associated with a Stp, which is also transmitted by P. leporinus and causes no differentiable symptoms in affected sugar beets (Gatineau et al., 2001, 2002; Sémétey et al., 2007a). In the present work the complex of planthopper cixiids involved with transmission of both Stp and SBRpr to sugar beet were studied.

## Materials and methods

Insect sampling was conducted in two sugar beet fields located in Jura department (France) during SBR epidemics, using transparent sticky traps maintained above the vegetation layer with stakes planted in the ground. Traps were posted at regular distances (10 meters) at nodes of a rectangular array in two sugar beet fields in 2005 (28

traps) and 2006 (72 traps). They were maintained in fields from the beginning of June until sugar beet harvesting, covering the flight time of most planthopper cixilds in temperate regions. Trapped cixilds were counted and removed every 2-3 days. Insects were preserved in 70% ethanol for later identification of uncertain species, or frozen at -80 °C for PCR detection of Stp and SBRpr.

Transmission assays were conducted with *C. wagneri*, *H. obsoletus* and *P. leporinus* collected in 2005 and 2006 with a D-Vac aspirator in sugar beet fields, on weeds and in fallow fields. *H. obsoletus* collected from bindweed (b) or nettle (n) were separated. Planthoppers were caged in groups of 4-5 (*C. wagneri* or *P. leporinus*), 8-10 (*H. obsoletus* b), or 20 (*H. obsoletus* n) specimens per plant, on 35-50 day-old healthy sugar beet seedlings. Unexposed seedlings were kept as healthy controls.

Stp and SBRp were identified by PCR and RFLP analysis using total DNA extracted from insects and plants. Insects were tested separately or in groups of 3 (*H. obsoletus*). 'Ca. Ph. fragariae' and SBRpr were identified by RFLP of a 16Sr DNA fragment amplified with primers Fra5 (Zreik et al., 1998) and rA\_PB\_2 (A. Bressan, unpublished.) Primer pairs Alb1/Oliv1 (Sémétey et al., 2007a) and Pfr1/Pfr4 (Foissac et al., 2000) were used for specific detection in insects of SBRpr and 'Ca. Ph. fragariae', respectively. Stp was detected with nested-PCR of Stol11 fragment (Clair et al., 2003). Stp in positive samples were typed according to Langer and Maixner (2004).

Parameters for virulence assessment of Stp and SBRpr were fresh weights of entire sugar beet and of tap root, and an estimation of sugar content by refractometric dried substance (RDS) of tap root pulp mesurement 5 months after planthopper pathogen transmission.

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**Table 1**. Infection of individual insects with SBR bacterium and two types of stolbur phytoplasma and their transmission efficiency in batches (see text) to sugar beet seedlings.

Planthopper species	SBR bacterium		Stolbur phytoplasma (bindweed)		Stolbur phytoplasma (nettle)	
	infected/tested	diseased/inoc.	infected/tested	diseased/inoc.	infected/tested	diseased/inoc.
	insects	sugar beets	insects	sugar beets	insects	sugar beets
P. leporinus	62/75	15/25	0/75	0/25	0/75	0/25
C. wagneri	38/83	12/25	0/83	0/25	0/83	0/25
H. obsoletus (b)	0/61*	0/19	38/61	13/19	0/61	0/19
H. obsoletus (n)	0/25*	0/36	0/25	0/36	19/25	0/36

<sup>\*</sup>H. obsoletus were tested in groups of three specimens.

#### **Results**

Insects - *P. leporinus* were much more abundant than *C. wagneri* or *H. obsoletus*. Of 4,033 and 10,047 cixiid specimens captured in 2005 and 2006 respectively, *P. leporinus* specimens represented 97.5% (3,933) and 98,7% (9,917), *C. wagneri* 1.9% (75) and 0.5% (51), and *H. obsoletus* 0.6% (25) and 0.8% (79), respectively. SBRpr was detected in *P. leporinus* and in *C. wagneri* but not in *H. obsoletus* (b) or (n) populations. None of 69 *C. wagneri* specimens were positive for '*Ca.* Ph. fragariae' (data not shown). Stp was not detected in either *P. leporinus* or *C. wagneri* whereas both (b) and (n) populations of *H. obsoletus* were phytoplasma positive (table 1). Stp types detected in *H. obsoletus* (b) or (n) were according to Langer and Maixner (2004).

Pathogens - *TaqI* restriction of Fra5/rA\_PB\_2 16S rDNA differentiate SBRpr from '*Ca*. Ph. fragariae' (not shown). SBRpr was transmitted by *P. leporinus* and *C. wagneri*, transmission of Stp was obtained only with *H. obsoletus* (b) (table 1) although survival on sugar beet of (b) and (n) populations was comparable (not shown).

Virulence on sugar beet - Similar symptoms and reduction of the biomass and sugar content were observed in plants inoculated with SBRpr or Stp(b). Data showed statistically significant differences from healthy controls, but the proportion of tap root weight to total plant weight was lower in Stp-infected than in SBRpr-infected sugar beets.

#### **Discussion**

The aboundant populations of *P. leporinus* in sugar beet fields with a very high proportion of SBRpr positive specimens, confirmed its major role in SBR disease spreading. Our data showed that virulence of Stp(b) to sugar beet was similar or higher than that of SBRpr. C. wagneri can also be a natural vector of SBRpr. We were not able to detect the Stp(n) type in sugar beets that remained symptomless although they were inoculated with phytoplasma positive H. obsoletus (n). In addition, it is possible that competition between Stp and SBRpr in plant tissues and insect organs may have progressively excluded the presence of Stp that was earlier detected in P. leporinus and sugar beets (Gatineau et al., 2001). This work underlines common traits between diseases associated with phytoplasmas and phloem-restricted proteobacteria and the new importance of latter diseases.

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### References

CLAIR D., LARRUE J., AUBERT G., GILLET J., CLOQUEMIN G. BOUDON-PADIEU E., 2003.- A multiplex nested-PCR assay for sensitive and simultaneous detection and direct identification of phytoplasma in the elm yellows group and stolbur group and its use in survey of grapevine yellows in France. *Vitis*, 42: 151-157.

DANET J.-L., FOISSAC X., ZREIK L., SALAR P., VERDIN E., NOURISSEAU J.-G., GARNIER M., 2002.- 'Candidatus Phlomobacter fragariae' is the prevalent agent of marginal chlorosis of strawberry in French production fields and is transmitted by the planthopper Cixius wagneri (China).- Phytopathology, 93: 644-649.

FOISSAC X., DANET J.-L., ZREIK L., GANDAR J., NOURRISSEAU J.-G., BOVÉ J.-M. GARNIER M., 2000.- Cloning of the *spoT* gene of '*Ca*. Phlomobacter fragariae' and development of a PCR-RFLP assay for detection of the bacterium in insects.-*Applied and Environmental Microbiology*, 66: 3474-3480.

GATINEAU F., LARRUE J., CLAIR D., LORTON F., RICHARD-MOLARD M., BOUDON-PADIEU E., 2001.- A new natural planthopper vector of stolbur phytoplasma in the genus *Pentastiridius* (Hemiptera: Cixiidae).- *European Journal of Plant Pathology*, 107: 263-271.

GATINEAU F., JACOB N., VAUTRIN S., LARRUE J., LHERMINIER J., RICHARD-MOLARD M; BOUDON-PADIEU E., 2002.-Association with the syndrome "basses richesses" of sugar beet of a phytoplasma and a bacterium-like organism transmitted by a *Pentastiridius* sp.- *Phytopathology*, 92: 384-392.

Langer M., Maixner M., 2007.- Molecular characterisation of grapevine yellows associated phytoplasmas of the stolburgroup based on RFLP-analysis of non ribosomal DNA.- *Vitis*, 43: 191-199.

SÉMÉTEY O., BRESSAN A., GATINEAU F., BOUDON-PADIEU E., 2007a.- Development with RISA of a specific assay for detection of the bacterial agent of the syndrome "basses richesses" of sugar beet.- *Plant Pathology*, in press.

SÉMÉTEY O., GATINEAU F., BRESSAN A., BOUDON-PADIEU E., 2007b.- Characterization of a γ-3 Proteobacteria responsible for the syndrome "basses richesses" of sugar beet transmitted by *Pentastiridius* sp. (Hemiptera: Cixiidae).- *Phytopathology*, 97: 72-78.

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