

# New observations of Coleoptera associated with Mantodea ootheca and an overview of the previous records

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## Abstract

This paper presents the first observations of the Coleoptera genera: *Attagenus* Latreille 1802 (Dermestidae), *Phradonoma* Jacquelin du Val 1859 (Dermestidae), and *Trichodes* Herbst 1792 (Cleridae) in association with Mantodea ootheca; in particular, it is the first time that predator/inquiline beetle larvae are recorded on *Bolivaria* Stal 1877 (Mantodea Rivetiniidae) ootheca. Furthermore, a new host species for *Thaumaglossa rufocapillata* Redtenbacher 1867 (Dermestidae) is documented. In addition, we provide a short review of the knowledge regarding the association between beetle larvae and Mantodea ootheca worldwide.

**Key words:** beetles, *Bolivaria*, Mantid, ootheca, larvae.

## Introduction

The ootheca is a protein-based structure produced by female mantids during oviposition which is meant to enclose and protect eggs from extreme environmental conditions, parasitoids, and predators (Hackman and Goldberg, 1960). Because of their protective function, multiple arthropods have evolved the advantageous ability to use both viable and evacuated ootheca as nesting sites or occasional shelters (Mirzaee *et al.*, 2021a; 2021b). Coleoptera has been recorded multiple times as inquiline and predators of mantis ootheca (Ramsay, 1990) and this type of association involves different beetle families and several different Mantodea genera worldwide (references cited in table 1). Predation and/or collateral activities by arthropods, including beetles, may compromise not only the structure and functionality of the ootheca but also affect eggs and nymphal survival rate, ultimately influencing the population dynamics of mantids in their habitats. This study aims to increase and update knowledge on interactions between Coleoptera and mantis ootheca, providing new species and new host records, together with a literature review on the subject.

## Materials and methods

### Specimens' collection

During research by the first author in 2020-2021 aimed at monitoring and documenting the biology of praying mantids of Iran, a series of ootheca were collected for more study in the laboratory. A total of twenty-eight ootheca belonging to *Bolivaria brachyptera* (Pallas 1773) (Mantodea Rivetiniidae), and *Hierodula tenuidentata* Saussure 1869 (Mantodea Mantidae) were collected in two districts of Fars (Darab: 28°46'45"N 54°34'08"E;

Dalkhan 30°14'32"N 52°06'09"E) and one district of Hormozgan (Bandar Abbas: 27°11'11"N 56°16'50"E) (figure 1) provinces. From these ootheca, 19 of them showed signs of predation or activities by other arthropods (four ootheca of *B. brachyptera*, and 15 of *H. tenuidentata*). Two ootheca of *B. brachyptera* were deposited on the rocks, four under rocks, and three were attached to the branches of thorny bushes. All *H. tenuidentata* ootheca were attached to tree branches, mostly at the middle (figure 2). They were removed from their deposition sites, placed in plastic jars with some cotton inside, and then stored under laboratory conditions (26 ± 1 °C, 40-50% RH, and 16L:8D). The ootheca were checked daily and their conditions recorded. The collected specimens were deposited in the Zoology Museum of Shiraz University, Shiraz- Iran (ZM-CBSU). Photos were taken using a Canon 700D digital camera.

Two ootheca of *Hierodula cf. patellifera* Serville 1839 (Mantodea Mantidae), a native species of the Japanese fauna, were collected in Chikusa-ku, Nagoya (Japan) by Shinichiro Ishikawa. The egg cases were conferred to Katsumi Akita, who stored them in sealed plastic cases until the emergence of both mantid nymphs and beetles. The specimens collected are deposited in the Katsumi Akita private collection, Tsu City, Mie Pref., Japan.

### Literature review

In order to compile and then review the most exhaustive literature on beetles associated with mantis ootheca (via Google Scholar), we used keywords such as "ootheca", "mantid\*", "beetle", "host", integrated using the Boolean operators AND, OR, NOT and the use of "" for specific word combinations. Further, we recovered information from volumes not available online or on PDF. The results of the research have been summarized in table 1.



**Figure 1.** General view of the collecting sites in Iran: **A)** Darab habitat, **B)** Dalkhan habitat, **C)** Bandar Abbas habitat.



**Figure 2.** **A)** Ootheca of *B. brachyptera* attached to the lower part of a stone; **B)** ootheca of *H. tenuidentata* on a tree branch.

**Table 1.** Recorded beetles associated with mantids oothecae from the world.

Beetle species	Beetle family	Mantid species	Country	Reference
<i>Opilo domesticus</i> (Sturm 1837)	Cleridae	<i>Sphodromantis viridis</i> (Forsk. 1775) (Mantidae)	Tunisia	Ehrmann, 2002
<i>Trichodes</i> sp.	Cleridae	<i>Bolivaria brachyptera</i> (Pallas 1773) (Rivetinidae)	Iran	this paper
<i>Anthrenocerus australis</i> (Hope 1843)	Dermestidae	<i>Miomantis</i> sp. (Miomantidae)	New Zealand?	Ramsay, 1990
<i>Anthrenus</i> sp.	Dermestidae	<i>Hierodula tenuidentata</i> Saussure 1869 (Mantidae)	Romania	Pintilioaie <i>et al.</i> , 2021
<i>Anthrenus</i> sp.	Dermestidae	<i>Stagnomantis Carolina</i> (Johansson 1763) (Mantidae)	USA	Rau and Rau, 1913
<i>Anthrenus (Nathrenus) verbasci</i> (L. 1767)	Dermestidae	<i>Hierodula tenuidentata</i> Saussure 1869 (Mantidae)	Romania	Rădac and Háva, 2021
<i>Attagenus fasciatus</i> (Thunberg 1795)	Dermestidae	<i>Bolivaria brachyptera</i> (Pallas 1773) (Rivetinidae)	Iran	this paper
<i>Dermestes maculatus</i> De Geer 1774	Dermestidae	<i>Archimantis latistyla</i> Serville 1839 (Mantidae)	Australia	Hawkeswood, 2003
<i>Dermestes</i> sp.	Dermestidae	<i>Hierodula saussurii</i> Kirby 1904 (Mantidae)	China	Kershaw, 1910
<i>Globicornis (Globicornis) nigripes</i> (F. 1792)	Dermestidae	<i>Hierodula tenuidentata</i> Saussure 1869 (Mantidae)	Romania	Rădac and Háva, 2021
<i>Orphinus</i> sp.	Dermestidae	<i>Archimantis latistyla</i> (Serville 1839) (Mantidae)	Australia	Coombs, 1994 a; 1994b
<i>Phradonoma nobile</i> (Reitter 1881)	Dermestidae	<i>Hierodula tenuidentata</i> Saussure 1869 (Mantidae)	Iran	this paper
<i>Thaumaglossa bimaculate</i> Arrow 1915	Dermestidae	<i>Sphodromantis gastrica</i> (Stal 1858) (Mantidae)	South Africa	Arrow, 1915
<i>Thaumaglossa bimaculate</i> Arrow 1915	Dermestidae	<i>Sphodromantis viridis</i> (Forsk. 1775) (Mantidae)	South Africa	Arrow, 1915
<i>Thaumaglossa indiana Indiana</i> Vijay Veer 2004	Dermestidae	<i>Hierodula westwoodi</i> Kirby 1904 (Mantidae)	India	Vijay Veer <i>et al.</i> , 2004
<i>Thaumaglossa indiana pakistana</i> Hava 2006	Dermestidae	<i>Hierodula westwoodi</i> Kirby 1904 (Mantidae)	Pakistan	Háva, 2006
<i>Thaumaglossa rufocapillata</i> Redtenbacher 1867	Dermestidae	<i>Tenoderangustipennis</i> Saussure 1869 (Mantidae)	Japan	Matsura, 1979
<i>Thaumaglossa hilleri</i> Reitter 1881	Dermestidae	<i>Tenodera aridifolia</i> Stoll 1813 (Mantidae)	Japan	Iwasati <i>et al.</i> , 2000
<i>Thaumaglossa hilleri</i> Reitter 1881	Dermestidae	<i>Tenoderangustipennis</i> Saussure 1869 (Mantidae)	Japan	Iwasati <i>et al.</i> , 2000
<i>Thaumaglossa rufocapillata</i> Redtenbacher 1867	Dermestidae	<i>Hierodula cf. patellifera</i> Serville 1839 (Mantidae)	Japan	this paper
<i>Thaumaglossa pauliani</i> Paulian 1953	Dermestidae	<i>Brancsikla</i> sp. (Majangidae)	Madagascar	Paulian, 1953
<i>Thaumaglossa pauliani</i> Paulian 1953	Dermestidae	<i>Empusa</i> sp. (Empusidae)	Madagascar	Paulian, 1953
<i>Thaumaglossa pauliani</i> Paulian 1953	Dermestidae	<i>Sphodromantis</i> sp. (Mantidae)	Madagascar	Paulian, 1953
<i>Thaumaglossa rufocapillata</i> Redtenbacher 1867	Dermestidae	<i>Tenodera</i> sp. (Mantidae)	Japan, Java	Iwasaki <i>et al.</i> , 1998; Dresner, 1970
<i>Thaumaglossa rufocapillata</i> Redtenbacher 1867	Dermestidae	<i>Tenodera aridifolia</i> Stoll 1813 (Mantidae)	Japan	Iwasaki <i>et al.</i> , 1996
<i>Thaumaglossa rufocapillata</i> Redtenbacher 1867	Dermestidae	<i>Tenoderangustipennis</i> Saussure 1869 (Mantidae)	Japan	Iwasaki <i>et al.</i> , 1996
<i>Thaumaglossa rufocapillata</i> Redtenbacher 1867	Dermestidae	Unknown	Indonesia	Dresener, 1970
<i>Thaumaglossa</i> sp.	Dermestidae	<i>Tenodera sinensis</i> (Saussure 1871) (Mantidae)	Japan	Curtis, 1940
<i>Thaumaglossa ooparasitica</i> Hava et Meriguet 2018	Dermestidae	<i>Paramantis prasina</i> (Serville 1839) (Mantidae)	Madagascar	Háva and Mériguet, 2018
<i>Thaumaglossa petrstanda</i> Hava 2003	Dermestidae	Unknown (Mantidae)	Bali	Háva and Suprayitno, 2020
<i>Trogoderma carteri</i> Armstrong 1942	Dermestidae	Unknown	Australia	Amstrong, 1949
<i>Trogoderma signatum</i> Sharp 1877	Dermestidae	<i>Miomantis</i> sp.? (Miomantidae)	New Zealand?	Ramsay, 1990
<i>Anthocomus (s.str.) fasciatus</i> (L. 1758)	Melyridae	<i>Hierodula tenuidentata</i> Saussure 1869 (Mantidae)	Romania	Plonski <i>et al.</i> , 2021
<i>Dasytes (Mesodasytes) aeratus</i> Stephens 1830	Melyridae	<i>Hierodula tenuidentata</i> Saussure 1869 (Mantidae)	Romania	Plonski <i>et al.</i> , 2021
<i>Dasytes</i> sp.	Melyridae	<i>Hierodula tenuidentata</i> Saussure 1869 (Mantidae)	Romania	Pintilioaie <i>et al.</i> , 2021
<i>Aplocnemus pectinatus</i> (Kuster 1849)	Rhadalidae	<i>Mantis religiosa</i> (L. 1758) (Mantidae)	Malta	Plonski <i>et al.</i> , 2021

## Results

The field observations and rearing of the specimens from the oothecae collected in Iran allowed the identification of the following beetles: *Attagenus fasciatus* (Thunberg 1795) (Dermestidae Attageninae), *Trichodes* sp. (Cleridae Trichodinae), and *Phradonoma nobile* (Reitter 1881) (Dermestidae Megatominae).

*A. fasciatus* was obtained from three oothecae of *B. brachyptera* (Mantodea Rivetiniidae) collected in Dalkhan (figure 3). These oothecae presented a few holes in the outer layer of the case; their removal from the substrate caused the exposure of the inner part that revealed the presence of beetle larvae. At the time of collection,

both viable eggs and mantid protonymphs were present. No nymphs emerged from the oothecae after the hatching of adult *Attagenus*, suggesting that their larvae completely fed upon mantid immature stages. All of these oothecae were 100% damaged by these beetles and no mantid nymphs emerged from them. *A. fasciatus* is a new faunistic record for the study area. This species was recorded by Modarres Awal (1997) from Tehran and also by Mroczkowski (1968), Zhantiev (1976), and Háva (2007; 2015) with no precise localities cited.

*Trichodes* sp. was recorded as larva when removing one ootheca of *B. brachyptera* that was attached to a stone (figure 4), in the Darab district. Before the discovery, the ootheca did not show any sign of attack; only the



**Figure 3.** *A. fasciatus*: **A)** mature larva, exuvia and immature adult collected from *B. brachyptera* ootheca; **B)** mature adult showing the typical colour pattern of the species.



**Figure 4.** *Trichodes* sp.: **A-B)** different views of the larva after the removal of the external part of the ootheca from the stone; **C)** dorsolateral view of the larva collected for the rearing under laboratory conditions.

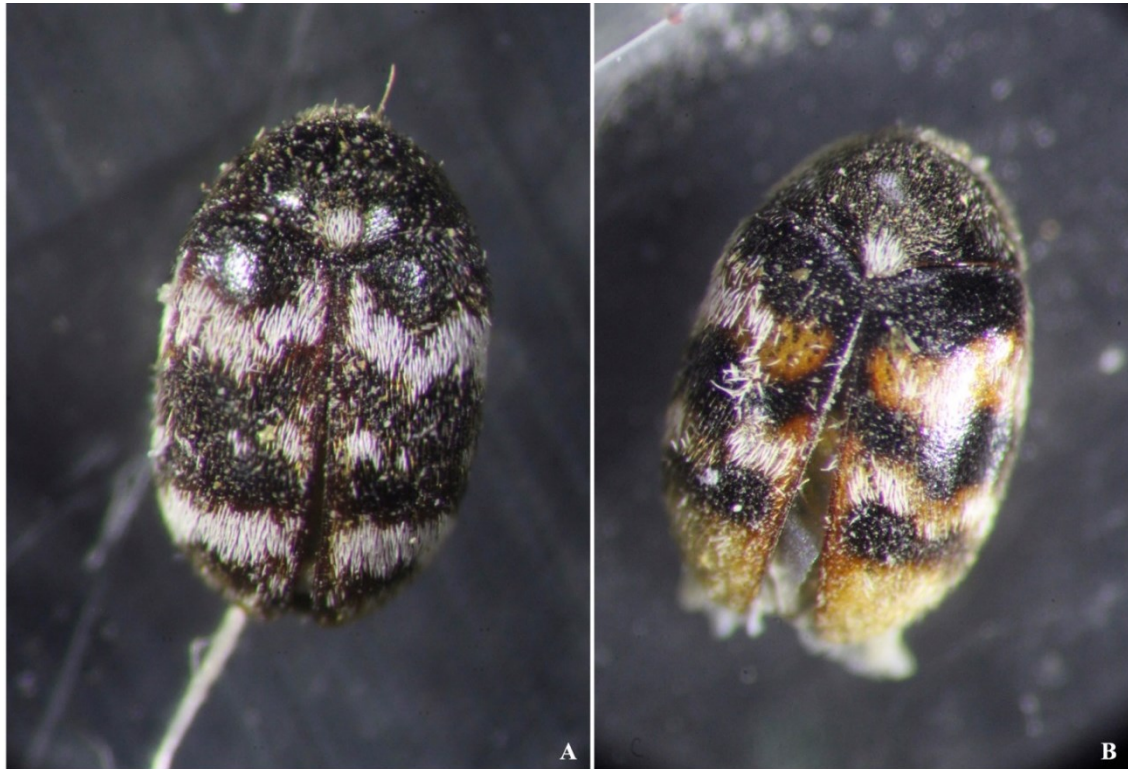
removal of the hardened cover showed how the beetle larva had mined the inner part and eaten all the eggs. Unfortunately, the rearing of the larva to the adult stage failed.

Both *Attagenus* and *Trichodes* were recorded in association with mantid oothecae for the first time; *B. brachyptera* represents a new host record.

*P. nobile* emerged from nineteen hatched oothecae of *H. tenuidentata* from the Bandar Abbas district (figure 5).

The adults hatched three months after the collection date. This record represents the first observation of the association between this species and the mantid oothecae.

In Japan, the rearing of *Hierodula* cf. *patellifera* under lab conditions resulted in 116 *Thaumaglossa rufocapillata* Redtenbacher 1867; it was interesting to note that the emergence of the beetles took place almost simultaneously with the emergence of the mantid nymphs (figure 6).



**Figure 5.** *P. nobile*: **A)** adult specimens showing the complete hairs vestiture on elytra; **B)** adult specimens lacking hair patches on elytra, showing the yellowish-orange spots on elytral integuments.



**Figure 6.** *T. rufocapillata*: **A)** adult beetles emerged in association with mantid nymphs; **B)** adult beetles remained on the hatched ootheca.

The literature review produced 25 records prior to the present contribution (table 1). Dermestidae, with 19 references, is the group most represented, followed by Melyridae (3), Cleridae (2), and Rhadalidae (1).

Dermestidae associated with mantid oothecae have been recorded in all the biogeographic regions, with the only exception of Antarctica and Neotropics, and include the genera *Dermestes* L. 1758 (Dermestinae Latreille 1807), *Anthrenocerus* Arrow 1915, *Anthrenus* Geoffroy 1762, *Globicornis* Latreille 1829, *Orphinus* Motschulsky 1858, *Thaumaglossa* Redtenbacher 1867 and *Trogoderma* Dejean 1821 (Megatominae Leach 1815). *Thaumaglossa* sp. and the species *Thaumaglossa indiana* Vijay Veer 2004, *Thaumaglossa pauliani* Paulian 1953 and *Thaumaglossa rufocapillata* Redtenbacher 1867 constitute the majority of records, further confirming the specialization of this genus for mantid oothecae, as indicated by Kiselyova and McHugh (2006).

Mantid genera that were involved in an association with Coleoptera larvae were primarily *Archimantis* Saussure 1869; *Hierodula* Burmeister 1838; *Mantis* L. 1758; *Sphodromantis* Stal 1871; *Stagmomantis* Saussure 1869; *Tenodera* Burmeister 1838; and then followed by Miomantidae, *Miomantis* Saussure 1870, and Empusidae, *Empusa* Illiger 1798.

## Discussion

According to the literature review and the new observations presented in the paper, mantid oothecae seem to be a valuable resource for several beetle larvae worldwide. It is interesting to note that oothecae of large-sized mantids (*Hierodula* spp. and *Tenodera* spp.) are preferred, possibly due to their greater size and volume that might offer more abundant trophic resources and a better shelter. The fact that several records involve native beetles in association with exotic mantid oothecae also suggests that the exploitation of the ootheca is generic and not species-specific.

The diversity of Dermestidae recorded on oothecae is attributable to their great ecological and trophic adaptability (Zhantiev, 2000; 2009); the larvae of this family are opportunistic scavengers, and capable of exploiting feeding substrates that vary in nutrients and water content (Ruzzier *et al.*, 2020; 2021). The dermestid subfamilies recorded, *i.e.*, Dermestinae, Attageninae Laporte 1840, and Megatominae, possess different levels of trophic specialization and it is plausible they develop at the expense of the oothecae at different ages. Dermestinae (*Dermestes* sp.) larvae specialize in feeding on substrates rich in water and are characterized by fast development, probably by predated fresh or relatively fresh oothecae, in the phases in which larvae and nymphs are still present. On the other hand, Attageninae (*Attagenus* sp.) and Megatominae (*Anthrenocerus* sp., *Anthrenus* spp., *Globicornis* sp., *Orphinus* sp., *Phradonoma* sp., *Thaumaglossa* spp., *Trogoderma* spp.), adapted to water-poor substrates and with slower larval development, might occur on mature oothecae and, especially the latter subfamily, may have

species capable of developing on very old oothecae, feeding on the ootheca itself or on insects remains, such as exuviae or dead inquiline arthropods. The biology of *Thaumaglossa* spp. (Megatominae) requires clarification as the only genus considered a specialist on mantid oothecae.

The occurrence of *Trichodes* sp. on mantid oothecae represents a novel observation for a genus that is usually a nest parasite of bees and wasps (Linsley and MacSwain, 1943; Carré, 1980). However, previous rare observations of larvae of *Trichodes* and other clerids feeding upon Orthoptera eggs (Arias *et al.*, 1994; Dysart, 2000) would seem to confirm that this family is able to parasitize orthopteroid insects and that is the current record might not just be a sporadic event.

The observations regarding Melyridae and Rhadalidae, as correctly argued by Plonski *et al.* (2021), constitute a possible case of predatory activity but most probably represent the use of the ootheca as a shelter for wintering.

Investigating these particular associations has many implications for species distribution dynamics. Recently, many mantids species have been recorded in non-native countries (see *e.g.* Schwarz and Ehrmann, 2018 for Europe or Andreson, 2018 for North America), mostly coming from Asia, including *H. tenuidentata* (Battiston *et al.*, 2018). The spread of these alien mantid species has been recorded as ootheca traveling attached to merchandise (Battiston *et al.*, 2020), sold for organic integrated pest control agents traditional medicine (Battiston *et al.*, 2010), or even released directly into the wild (Shcherbakov and Govorov, 2020) presumably along with alien parasites that may have an impact on native mantids species, on other fauna, or even have an economic impact on food supplies as pests.

## Conclusions

Mantis oothecae undoubtedly represent an important resource for various groups of Coleoptera, both for food and protection purposes. However, the true nature of these interactions remains, at least partly, obscure especially given the difficulty of collecting data and conducting observations in a systematic way. In particular, further investigations are needed in order to qualify whether beetles can be considered predators or parasites of mantids immature stages or inquiline species on oothecae.

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